

Alexander A. Vinks · Hartmut Derendorf
Johan W. Mouton *Editors*

Fundamentals of Antimicrobial Pharmacokinetics and Pharmacodynamics

 Springer

Fundamentals of Antimicrobial Pharmacokinetics and Pharmacodynamics

Alexander A. Vinks • Hartmut Derendorf
Johan W. Mouton
Editors

Fundamentals of Antimicrobial Pharmacokinetics and Pharmacodynamics

 Springer

Editors

Alexander A. Vinks
Division of Clinical Pharmacology
Cincinnati Children's Hospital
Medical Center and Department of
Pediatrics
University of Cincinnati
College of Medicine
Cincinnati, OH, USA

Hartmut Derendorf
Department of Pharmaceutics
University of Florida
Gainesville College of Pharmacy
Gainesville, FL, USA

Johan W. Mouton
Department of Medical Microbiology
Radboudumc, Radboud University Nijmegen
Nijmegen, The Netherlands

ISBN 978-0-387-75612-7 ISBN 978-0-387-75613-4 (eBook)

DOI 10.1007/978-0-387-75613-4

Springer New York Heidelberg Dordrecht London

Library of Congress Control Number: 2013953328

© Springer Science+Business Media New York 2014

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed. Exempted from this legal reservation are brief excerpts in connection with reviews or scholarly analysis or material supplied specifically for the purpose of being entered and executed on a computer system, for exclusive use by the purchaser of the work. Duplication of this publication or parts thereof is permitted only under the provisions of the Copyright Law of the Publisher's location, in its current version, and permission for use must always be obtained from Springer. Permissions for use may be obtained through RightsLink at the Copyright Clearance Center. Violations are liable to prosecution under the respective Copyright Law.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

While the advice and information in this book are believed to be true and accurate at the date of publication, neither the authors nor the editors nor the publisher can accept any legal responsibility for any errors or omissions that may be made. The publisher makes no warranty, express or implied, with respect to the material contained herein.

Printed on acid-free paper

Springer is part of Springer Science+Business Media (www.springer.com)

Contents

Part I Basic Concepts and Principles

1 Introduction to Pharmacodynamics	3
William A. Craig	
2 In Vitro and Animal PK/PD Models	23
William A. Craig	
3 Setting Clinical MIC Breakpoints from a PK/PD Point of View: It Is the Dose That Matters	45
Johan W. Mouton	
4 Principles of Applied Pharmacokinetic–Pharmacodynamic Modeling	63
Benjamin Wu, Sherwin K.B. Sy, and Hartmut Derendorf	
5 Pharmacodynamic In Vitro Models to Determine the Effect of Antibiotics	81
Julia Michael, Aline Barth, Charlotte Kloft, and Hartmut Derendorf	
6 Population Pharmacokinetic–Pharmacodynamic (PK/PD) Modeling of Anti-infective Agents and Its Applications to Individualized Therapy	113
Alexander A. Vinks	
7 Suppressing Resistance Development	135
Vincent H. Tam	
8 Drug–Drug Combinations	153
John Turnidge	

Part II Clinically Oriented Chapters

9 Aminoglycosides	201
Catharine C. Bulik, Charles H. Nightingale, and David P. Nicolau	
10 Continuous Infusion of Beta-lactam Antibiotics	223
Anouk E. Muller and Johan W. Mouton	
11 Macrolides and Ketolides	257
Françoise Van Bambeke	
12 Glycopeptides	279
Inge C. Gyssens	
13 Clinical Pharmacodynamics of Quinolones	323
George L. Drusano, H.S. Heine, and A. Louie	
14 Pharmacokinetics and Pharmacodynamics of Colistin	351
Roger L. Nation, Phillip J. Bergen, and Jian Li	
15 Daptomycin: Pharmacokinetic, Pharmacodynamic, and Dose Optimization	381
Céline Vidaillac and Michael J. Rybak	
16 PK/PD of Oxazolidinones	401
Ursula Theuretzbacher	
17 Tigecycline	445
Catharine C. Bulik, Anthony M. Nicasio, and Paul G. Ambrose	
Index	457

Contributors

Paul G. Ambrose, Pharm.D., F.I.D.S.A. Institute for Clinical Pharmacodynamics, Latham, NY, USA

Françoise Van Bambeke, Pharm.D., Ph.D. Louvain Drug Research Institute, Université catholique de Louvain, Pharmacologie cellulaire et moléculaire, Brussels, Belgium

Aline Barth, M.S. Department of Pharmaceutics, University of Florida, Gainesville, FL, USA

Phillip J. Bergen, Ph.D. Centre for Medicine Use and Safety and Monash Institute of Pharmaceutical Sciences, Monash University, Melbourne, VIC, Australia

Catharine C. Bulik, Pharm.D. Institute for Clinical Pharmacodynamics, Latham, NY, USA

William A. Craig, M.D. Division of Infectious Disease, University of Wisconsin School of Medicine and Public Health, Madison, WI, USA

Hartmut Derendorf, Ph.D. Department of Pharmaceutics, University of Florida, Gainesville, FL, USA

George L. Drusano, M.D. Institute for Therapeutic Innovation, University of Florida College of Medicine, Lake Nona, FL, USA

Inge C. Gyssens, M.D., Ph.D. Nijmegen Institute for Infection, Inflammation, and Immunity and Department of Medicine, Radboud University Medical Center, Nijmegen, The Netherlands

Department of Medical Microbiology and Infectious Diseases, Canisius Wilhelmina Hospital, Nijmegen, The Netherlands
Hasselt University, Hasselt, Belgium

Charlotte Kloft, Ph.D. Department of Clinical Pharmacy & Biochemistry, Freie Universitaet Berlin, Berlin, Germany

Julia Michael, Ph.D. Department of Clinical Pharmacy, Martin-Luther-Universitaet Halle-Wittenberg, Berlin, Germany

Johan W. Mouton, M.D., Ph.D., F.I.D.S.A. Department of Medical Microbiology, Radboudumc, Radboud University Nijmegen, Nijmegen, The Netherlands

Anouk E. Muller, M.D., Ph.D. Department of Medical Microbiology, Netherlands

Roger L. Nation, Ph.D. Monash Institute of Pharmaceutical Sciences, Monash University, Melbourne, Victoria, Australia

Anthony M. Nicasio, Pharm.D. Albany College of Pharmacy and Health Sciences, Albany, NY, USA

David P. Nicolau, Pharm.D., F.C.C.P., F.I.D.S.A. Division of Infectious Diseases and Pharmacy, Departments of Medicine, Center for Anti-Infective Research and Development, Harford Hospital, Hartford, CT, USA

Charles H. Nightingale, Ph.D. Center for Anti-Infective Research and Development, Harford Hospital, Hartford, CT, USA

Jian Li, Ph.D. Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Melbourne, VIC, Australia

Michael J. Rybak, Pharm.D., M.P.H. Anti-Infective Research Laboratory, Pharmacy Practice – 4148, Eugene Applebaum College of Pharmacy and Health Sciences, Wayne State University, Detroit, MI, USA

Sherwin K.B. Sy, M.S. Department of Pharmaceutics, University of Florida, Gainesville, FL, USA

Vincent H. Tam, Pharm.D. Department of Clinical Sciences and Administration, University of Houston College of Pharmacy, Houston, TX, USA

Ursula Theuretzbacher, Ph.D. Center for Anti-Infective Ages, Vienna, Austria

John Turnidge, M.B., B.S., F.R.A.C.P., F.R.C.P.A., F.A.S.M. Department of Pathology, University of Adelaide and SA Pathology, Women's and Children's Hospital, North Adelaide, South Australia, Australia

Department of Pediatrics, University of Adelaide and SA Pathology, Women's and Children's Hospital, North Adelaide, South Australia, Australia

Department of Molecular and Biomedical Science, University of Adelaide and SA Pathology, Women's and Children's Hospital, North Adelaide, South Australia, Australia

Céline Vidaillac, Pharm.D., Ph.D. Anti-Infective Research Laboratory, Pharmacy Practice – 4148, Eugene Applebaum College of Pharmacy and Health Sciences, Wayne State University, Detroit, MI, USA

Alexander A. Vinks, Pharm.D, Ph.D., F.C.P. Division of Clinical Pharmacology, Cincinnati Children's Hospital Medical Center and Department of Pediatrics, University of Cincinnati College of Medicine, Cincinnati, OH, USA

Benjamin Wu, Ph.D. Pharmacokinetic and Drug Metabolism, Amgen Inc., Thousand Oaks, CA, USA

Part I
Basic Concepts and Principles

Chapter 1

Introduction to Pharmacodynamics

William A. Craig

Abstract Since the early appreciation of differences in the time course of antimicrobial activity, much has been learned about the pharmacodynamics of antimicrobials. Specific PK/PD indices have been identified which are of major importance for efficacy and for the prevention of the emergence of resistance. Of major importance, the magnitudes of these PK/PD indices for efficacy have been shown to be very similar in animal infection models and human infections. Modeling has also identified that there are few differences in the index magnitude with different dosing intervals, among drugs within the same antimicrobial class (providing free drug concentrations are used), with different infection sites (except occasionally for pneumonia), and among susceptible and resistant strains of the same type of bacteria. Addition studies have shown that the magnitude of indices can increase significantly with a higher inoculum for *S. aureus* and that neutrophils have a minor enhancing effect on antimicrobial activity against Enterobacteriaceae but a more variable enhancing effect on activity against *S. pneumoniae* for different antimicrobials. Pharmacodynamic modeling has many applications including establishing new optimal dosing regimens, developing new antimicrobials and formulations, determining susceptibility breakpoints, providing guidelines for empiric therapy, and formulary development.

Keywords Pharmacodynamics • Modeling • PK/PD indices • Protein binding • Animal models • Neutrophil activity • Inoculum size • Susceptibility testing

W.A. Craig, M.D. (✉)

Division of Infectious Disease, University of Wisconsin School of Medicine and Public Health, MFCB-5th Floor, 1685 Highland Avenue, Madison, WI 53705-2281, USA
e-mail: wac@medicine.wisc.edu

Introduction

Antimicrobial pharmacodynamics deals with the relationship between measures of drug exposure and the efficacy and toxicity of antimicrobial agents. Since the early days of penicillin, researchers have been interested in determining which pharmacokinetic parameter is most important in determining microbiologic and clinical efficacy. For example, bacterial killing of staphylococci by penicillin was much different than by streptomycin (Garrod 1948). The rate of killing by penicillin was not dependent on the height of the drug concentration, while streptomycin demonstrated enhanced killing at higher concentrations. Studies in mice-infection models suggested that the duration of drug exposure was the most important parameter determining in vivo therapeutic efficacy of penicillin (Eagle et al. 1950). Interest in antimicrobial pharmacodynamics increased in the 1960s and 1970s when infections due to *Pseudomonas aeruginosa* with high MICs to multiple drugs appeared with increasing frequency (Rolinson 1973). This interest in antimicrobial pharmacodynamics has been further enhanced by the emergence of antimicrobial resistance to many drugs during the last 15–20 years.

Time Course of Antimicrobial Activity

A major determinant of the time course of antimicrobial activity is whether the drug exhibits bactericidal activity and whether the killing is enhanced by increasing concentrations or by longer exposure times. The second major determinant is whether the drug exhibits persistent inhibition of growth that lasts after the drug exposure. There are numerous in vitro persistent effects described in the literature that usually act together in the in vivo situation. The in vitro postantibiotic effect (PAE) describes the extent of continuing retardation in organism growth when the drug is suddenly removed by repeated washing, dilution, filtration, or inactivation (McDonald et al. 1977; Bundtzen et al. 1981). The postantibiotic sub-MIC effect (PA-SME) identifies additional prolongation in regrowth that results from sub-MIC drug concentrations (Cars and Odenholt-Tornqvist 1993). The postantibiotic leukocyte enhancement (PALE) identifies growth retardation that occurs when organisms in the postantibiotic state of growth are exposed to leukocytes (McDonald et al. 1981).

As stated above, these various in vitro persistent effects act together in describing the in vivo activity of antimicrobials. By injecting penicillinase intravenously in neutropenic mice when drug levels of piperacillin or aspoxicillin were expected to drop below the MIC, Oshida et al. (1990) were able to show that sub-MIC concentrations accounted for a little less than half of the 3.3 and 5.2 h in vivo postantibiotic effect observed with both drugs, respectively, against *Staphylococcus aureus*. Increasing the dose (and AUC) of amikacin sixfold increased the duration of the in vivo postantibiotic effect with *Klebsiella pneumoniae* in neutropenic mice from 3.4 to 7.4 h, while the duration of sub-MIC values was less than an hour and virtually the same for both doses (Craig et al. 1991). However, prolonging the half-life of

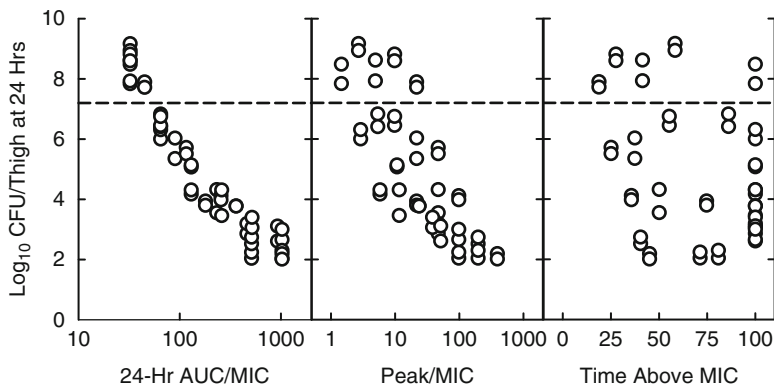


Fig. 1.1 Relationship between three PK/PD indices for total drug of levofloxacin and the \log_{10} CFU/thigh at 24 h for *Streptococcus pneumoniae* ATCC 10813 in the thighs of neutropenic mice. Reproduced with permission from Andes and Craig (2002)

amikacin from 18 to 110 min by inducing renal impairment also enhanced the AUC about sixfold, but the longer duration of sub-MIC concentrations increased the in vivo postantibiotic effect from 7.4 to 12.2 h. The role of leukocytes on the in vivo PAE has also been assessed. Studies with similar doses of gentamicin against the same strain of *K. pneumoniae* have reported in vivo PAEs of 7.8, 12.0, and 16.5 h in neutropenic, normal, and granulocytic mice, respectively (Shimizu et al. 1989).

Patterns of Antimicrobial Activity

Three major patterns of antimicrobial activity have been observed. The first applies to antimicrobials with concentration-dependent killing along with prolonged persistent effects. This pattern is observed with aminoglycosides, fluoroquinolones, polymyxins, daptomycin, and some of the new glycopeptides, such as telavancin and oritavancin, which also exhibit an additional membrane effect mechanism of action. One would predict that the ratio of the AUC and peak concentration to the MIC would be the primary PK/PD indices correlating with antimicrobial efficacy. Done-fractionation studies in animal models of infection in which five or six total doses are divided into many smaller doses given at different dosing frequencies have been useful in reducing the interdependence among the PK/PD indices and confirming which PK/PD index is most important for efficacy. The relationship of all the PK/PD indices based on total drug concentrations (protein binding in mice 15 %) to efficacy of levofloxacin against *Streptococcus pneumoniae* in the thighs of neutropenic mice are shown in Fig. 1.1 (Andes and Craig 2002). The 24-h AUC/MIC showed the best correlation for efficacy followed by the peak/MIC ratio. The time above MIC looked more like a scattergram.

The second pattern of antimicrobial activity is the exact opposite of the first pattern with concentration-independent killing and no or very short persistent effects. This pattern is characteristic of all of the β -lactam antibiotics, such as penicillins,

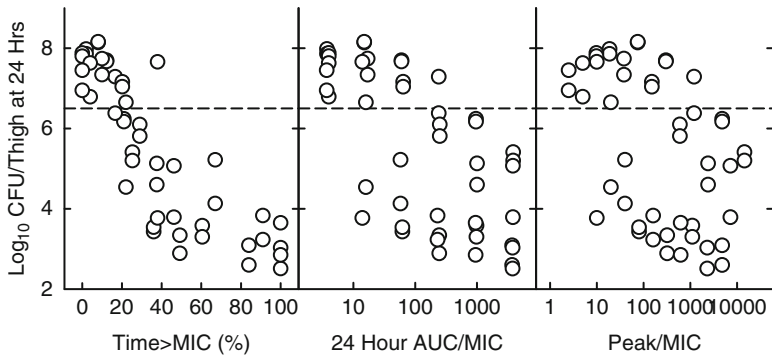


Fig. 1.2 Relationship between three PK/PD indices for total drug of imipenem and the \log_{10} CFU/thigh over 24 h for *Pseudomonas aeruginosa* ATCC 27853 in the thighs of neutropenic mice

cephalosporins, carbapenems, and monobactams. With this pattern, one would predict that the duration of time that active antibiotic concentrations exceeded the MIC would be the important PK/PD index for efficacy. Figure 1.2 demonstrates the relationships among the various PK/PD indices for total drug concentration of imipenem, a carbapenem β -lactam antibiotic with protein binding $<5\%$ in mice, against a standard strain of *Pseudomonas aeruginosa* in the thighs of neutropenic mice. The percentage of the dosing interval that concentrations exceeded the MIC showed the best correlation with organism growth and killing, while the relationships with AUC/MIC and peak/MIC looked more like scattergrams.

The third pattern of antimicrobial activity also exhibits concentration-independent killing but these antimicrobials induce prolonged persistent effects. This pattern is observed with a large number of antimicrobials including the tetracyclines, tigecycline, macrolides, azithromycin, clindamycin, linezolid and other oxazolidinones, chloramphenicol, trimethoprim, sulfonamides, vancomycin, and dalbavancin. Because the prolonged persistent effects will protect against regrowth when active drug concentration fall below the MIC, one would predict that the amount of drug or the AUC/MIC would be the important PK/PD index for these drugs. Figure 1.3 illustrates that relationship between the change in efficacy from the start of therapy and the various PK/PD indices based on total drug concentrations for vancomycin (protein binding 13% in mice) (Rybak 2006). The best correlation for efficacy was seen with 24-h AUC/MIC index. Peak/MIC and time above MIC showed much more variation in efficacy at different magnitudes of the index.

Magnitude of Index Required for Efficacy

Once the important PK/PD index driving efficacy is identified, the next piece of information needed is what magnitude of the index is required for antimicrobial efficacy. A large number of animal studies on the efficacy of β -lactams against